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Dear Dr. Migliore:

Thanks for your letter dated May 3, 1995. Please find enclosed a summary of the progress report on my Moran Foundation project entitled 'Cloning and sequence analysis of the mouse  $\gamma$ glutamylcysteine synthetase ( $\gamma$ GCS) gene'. During the past year my efforts has been directed towards characterization of the genomic structure of the mouse  $\gamma$ GCS gene. I am happy to inform you that I have completed sequencing and characterizing about 90% of the mouse  $\gamma$ GCS gene and will start preparing a manuscript for publication in the very near future.

I am also in the process of writing a new but related research proposal entitled "Disruption of the  $\gamma$ glutamylcysteine synthetase ( $\gamma$ GCS) locus in embryo-derived stem (ES) cells" which I will submit to The Moran Foundation for financial consideration for the fiscal year 1995-1996.

I would like to thank the Scientific Advisory Committee and the Moran Foundation for supporting this research.

Sincerely yours,

*Joseph Osei-Frimpong, Ph.D.*  
Joseph Osei-Frimpong, Ph.D.

**Progress Report - Cloning and sequence analysis of the of the mouse  $\gamma$ glutamylcysteine synthetase ( $\gamma$ GCS) gene - [3-93-0067]**

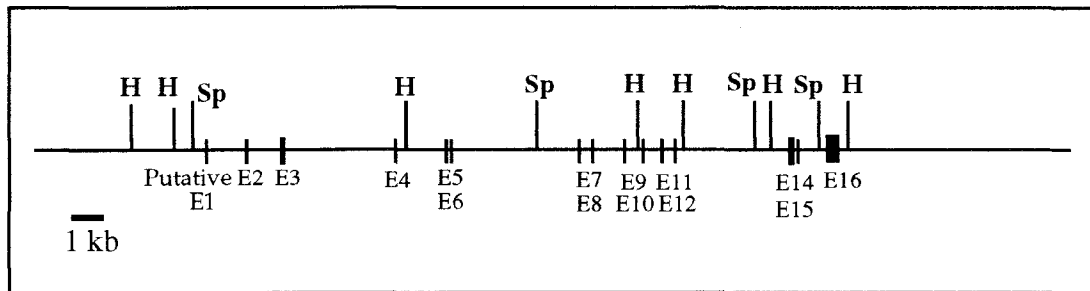
**Characterization of the Genomic Structure of the Mouse  $\gamma$ GCS Gene**

**A. Cloning of mouse  $\gamma$ GCS cDNA**

The  $\gamma$ GCS  $\lambda$ -phage clones characterized previously (see last year's Progress Report) were originally isolated using a rat  $\gamma$ GCS cDNA probe. Identification of mouse  $\gamma$ GCS intron-exon borders and 5'-flanking regions would be greatly facilitated by access to mouse  $\gamma$ GCS cDNA clones, since cDNAs from different species are most likely to diverge at the 5' and 3' non-coding regions. Using a modification of the rapid amplification of 5'-cDNA ends (5'-RACE) procedure, I have cloned a partial mouse  $\gamma$ GCS cDNA (the first 300 bp from the ATG) from mouse kidney. This clone was sequenced and found to have a 94% sequence similarity index with the same region of the rat cDNA.

**B. Exonic Structure of the mouse  $\gamma$ GCS Gene**

Using Southern blotting with mouse  $\gamma$ GCS cDNA probes, high speed sequencing and PCR (for introns > 300 bp) I have deduced the exonic structure of the mouse gene at the 5'-most coding region from my  $\gamma$ GCS  $\lambda$  clones. Also based on published rat cDNA sequence (pending the cloning of a full length mouse cDNA), I have also tentatively deduced the rest of the exonic structure of the mouse gene, with the exception of exons one and thirteen, which are yet to be precisely localized and sequenced. All exons are contained on the tentative map deduced from alignment of the  $\gamma$ GCS clones (see Progress Report of last year). This tentative genomic structure of the mouse  $\gamma$ GCS gene (Osei-Frimpong, Lebovitz, Lieberman, unpublished) is shown in Figure 1 below. The mouse  $\gamma$ GCS heavy subunit gene most likely consists of 16 exons and spans at least 23 kb. The biggest exon is the 16 th exon (465 bp), which consists of both coding and non-coding regions. The smallest exon is the 5 th exon (59 bp).



**Figure 1. Genomic structure of the mouse  $\gamma$ GCS heavy subunit gene (unpublished); exons are represented by black boxes. All exon and intron lengths are drawn to scale. H-HindIII, Sp-SpeI, E-exon, kb-kilobase**