

TITLE: APPLICATIONS OF HPLC AMINO ACID DETERMINATIONS (1-85-0013)

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Over the past year, we have used Moran Foundation funds for the measurement of serum amino acids in head injured patients using high pressure liquid chromatography with post-column ninhydrin derivatization. Though technically demanding, this technique permits accurate detection of some 40 serum constituents including all the essential and nonessential amino acids.

Pickering Laboratories of Mountain View, California, was the source of columns and reagents for a post-column ninhydrin derivatization method of measuring amino acids in physiological fluids. Chromatographic conditions included a 3mm X 250mm 10 $\mu$ M lithium cation exchange column at 42°C with a guard column and a three component lithium salt mobile phase (flow rate 0.3 ml/min) gradient with an elution and regeneration cycle of 210 minutes (figure 1). Following separation, the amino acids were mixed with ninhydrin (flow rate 0.3 ml/min) at a mixing tee and subsequently reacted for 100 seconds at 120°C in a Pickering CRX390 post column reactor resulting in chromophores monitored at 546 nm in a 12 $\mu$ l cell (figure 2). An injection of standard and a serum sample on the system in its current configuration are shown (figures 3 & 4). Sample preparation involved simple deproteinization followed by autoinjection of a 20  $\mu$ l sample.

We have injected 20 standards and 80 serum samples with good preservation of constituent resolution and peak areas. At this point, we

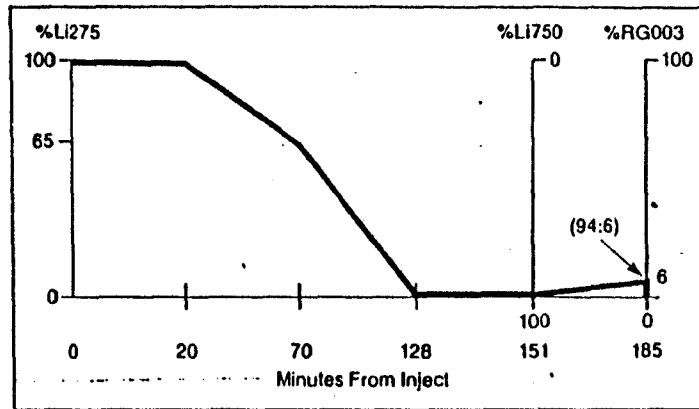
have about 20 more serum samples to process through the first week in December 1985 as well as the analysis of standards containing a few substances not contained in our original standard solution, but which are present and of interest in serum. Analysis of all of the data cannot be completed until these additional standards are used, but we anticipate completion of all data analysis by the end of December 1985. We have demonstrated already, however, that gamma-aminobutyric acid is not elevated in comatose head injured patients as was suggested by preliminary experiments in which the amino acid analyses were performed outside of the medical center.

We have demonstrated that this method can be implemented in a clinical laboratory with gradient high pressure liquid chromatographic capability. The pre-mixed reagents are reasonably inexpensive further facilitating service laboratory implementation. A major drawback of the method is the analysis time of 210 minutes which limits the total number of samples which can be processed to about 7/day. We hope to complete our data analysis in the next few weeks in time for submission of abstracts to national clinical chemistry and critical care meetings. Additionally, the data from the amino acid experiment will be linked to cerebral metabolic and systemic nitrogen balance data we have on the patients in the study.

SUMMARY: The serum amino acid profile in metabolic disturbances following head injury and other critical illnesses is not used widely in clinical practice in part due to the lack of rapid, reliable methods for measuring amino acids in physiological fluids which could be implemented in service clinical laboratories. Recent developments in chromatographic technology described here may now permit the amino acid analyses to be

performed in a matter of hours in any clinical laboratory with competence in high performance liquid chromatography.

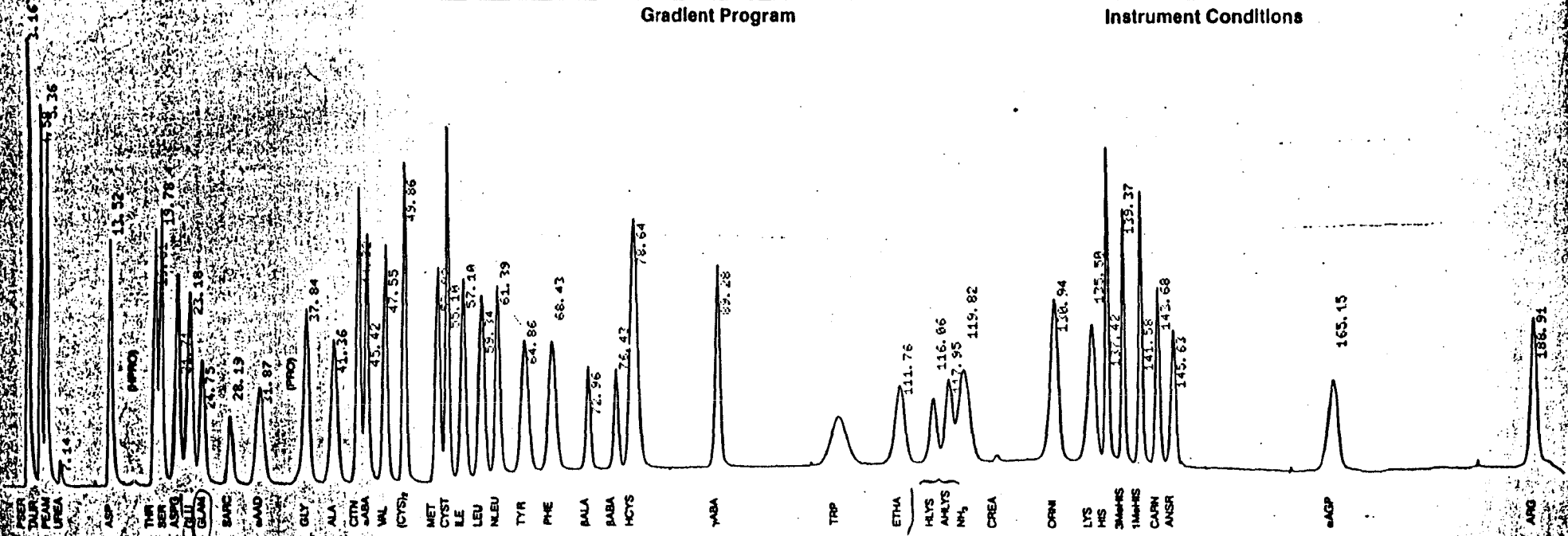
# GRADIENT LC, AMINO ACIDS: Physiologic Fluid



Gradient Program

**Sample:** 2.5 nM/Constituent  
**Flow Rate:** 0.3cc/min, Eluent  
 0.3cc/min, Trione  
**Temperature:** 42°C Isothermal  
**Development:** 50 sec at 130°C  
**Detector:** 0.5 AUFS  
 25µL x 12mm

Instrument Conditions

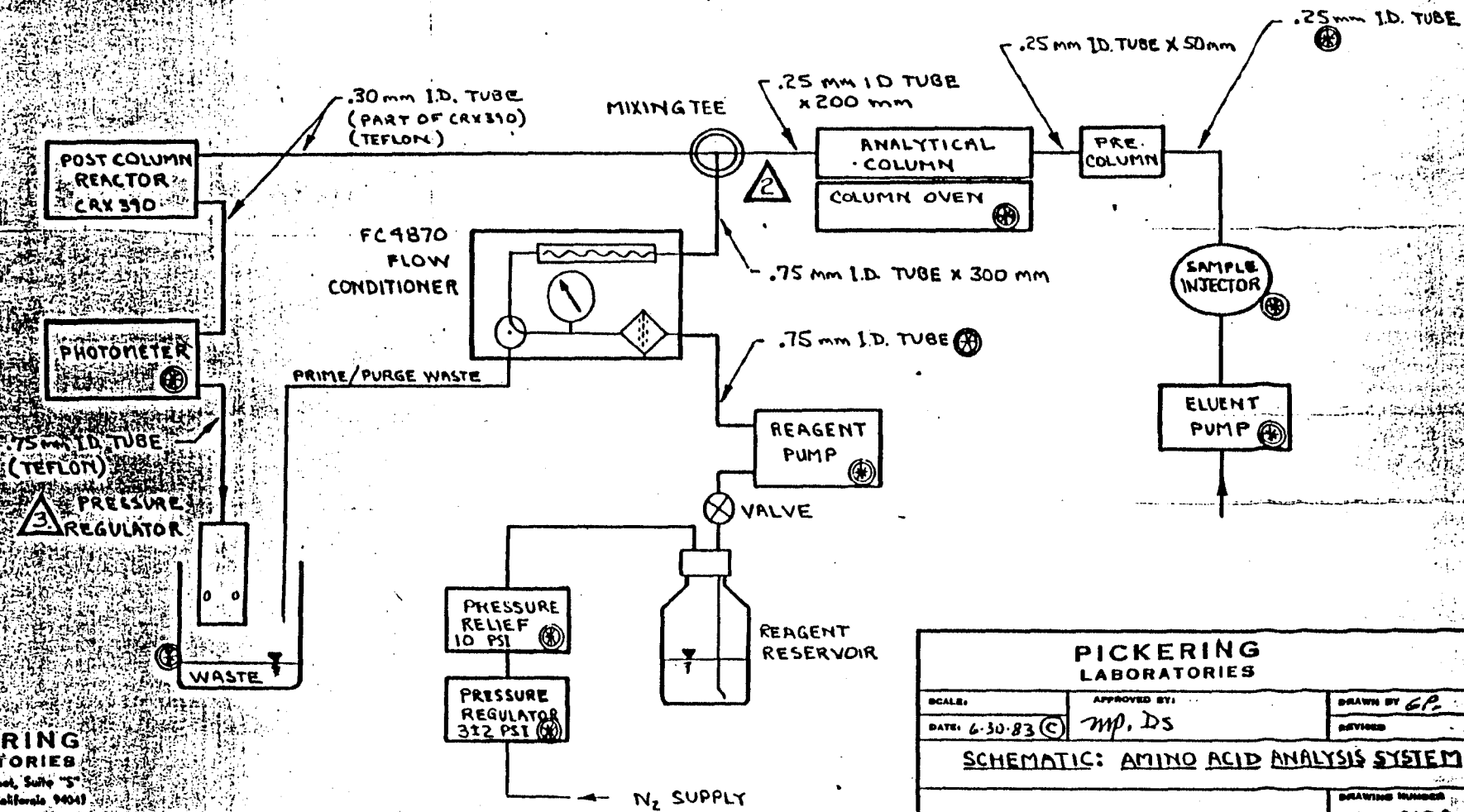


**PICKERING  
LABORATORIES**

NOTES: 1. (⊗) ITEMS NOT SUPPLIED BY PICKERING LABORATORIES.

2. **WARNING:** DO NOT PERMIT REAGENT TO BACKFLOW INTO ANALYTICAL COLUMN. REAGENT WILL DAMAGE COLUMN.

3. **WARNING:** REAGENT AND ELUENT BOIL AT 100°C. REACTOR TUBE WILL PLUG AND BURST IF OPERATED ABOVE 100°C WITHOUT SPECIFIED REGULATED BACK-PRESSURE.

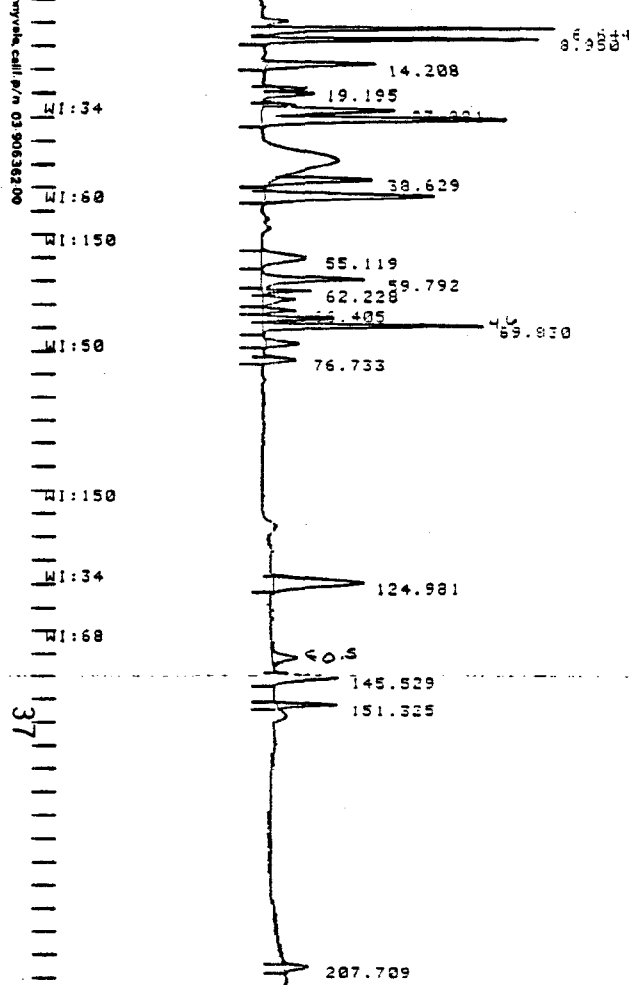


PICKERING  
LABORATORIES  
1951 Colony Street, Suite "5"  
Mountain View, California 94041

<b>PICKERING LABORATORIES</b>		
SCALE:	APPROVED BY: <i>mp, ds</i>	DRAWN BY: <i>GP</i>
DATE: 6-30-83 ©		REVISED:
<b>SCHEMATIC: AMINO ACID ANALYSIS SYSTEM</b>		
		DRAWING NUMBER: <b>1100-0180</b>



CHART SPEED 0.1 CM/MIN  
ATTEN: 256 ZERO: 10% 5 MIN/TICK



RECALC  
TITLE: AMINO ACID ANALYSIS

11:42 20 JUL 85

CHANNEL NO: 1

SAMPLE: 6772A

METHOD: AA

PEAK NO	PEAK NAME	RESULT UM/L	TIME (MIN)	TIME OFFSET	AREA COUNTS	SEP CODE	WI/2 (SEC)
1	TAUR	285.8	6.641	-0.000	3157030	BB	32.00
2	UREA	4233.8	8.950	-0.000	11897610	BB	50.15
3		166.8	14.208		20000000	BB	45.40
4		310.8	19.195		37091000	BB	
5	SER	337.8	22.551	-0.001	24484000	BB	32.20
6	ASPG	1583.8	25.983	-0.000	22031000	VV	50.10
7	GLU	1483.1	29.988	-0.000	47170000	VV	55.85
8	PRO	1489.8	35.629	-0.000	18571000	BB	55.85
9	ALA	225.8	42.161	-1.100	38852500	BB	70.45
10	VAL	138.1	55.119	-1.101	15771400	BB	100.00
11		178.8	59.792		21504000	VV	55.50
12	(CYS)2	33.8	62.228	-0.000	50900000	VV	24.90
13	MET	55.8	63.991	-0.018	54635900	VV	52.60
14	ILEU	35.8	66.405	-0.195	42684200	VV	38.15
15	LEU	80.8	68.120	-0.170	36197900	VV	39.70
16	NLEU	INT STD	69.830	-0.000	30061100	VV	40.05
17	TYR	54.2	73.383	-0.007	62892500	BB	55.80
18	PHE	52.8	76.733	0.063	60496800	BB	55.05
19	HLYS	163.8	124.981	0.071	29809000	BB	71.75
20	LYS	112.2	145.529	0.045	16344600	BB	71.40
21	HIS	60.6	151.325	0.105	80227900	BB	40.15
22	ARG	50.2	207.709	0.585	56724100	BB	71.50
	ORN?	34.9					
TOTALS:		8256.2		-1.131	38710000		

DETECTED PKS: 22 REJECTED PKS: 0

DIVISOR: 1.00000 AMT STD: 250.000 MULTIPLIER: 1.00000

NOISE: 110.5 OFFSET: 8028